

Pathogenesis and Experimental Models of Cerebral Malaria: A Review

Prati Pal Singh and
Bhanu Prakash

Centre of Infectious Diseases, Department of Pharmacology and Toxicology, National Institute of Pharmaceutical Education and Research, SAS Nagar, 160 062, Punjab, India

***Corresponding author:**

Prof. Prati Pal Singh

✉ drppsingh2016@gmail.com

Centre of Infectious Diseases, Department of Pharmacology and Toxicology, National Institute of Pharmaceutical Education and Research, SAS Nagar, 160 062, Punjab, India.

Citation: Singh PP, Prakash B (2018) Pathogenesis and Experimental Models of Cerebral Malaria: A Review. J Pharm Microbiol. Vol. 4 No. 1:3

Abstract

Cerebral malaria (CM), a severe complication of the nervous system due to uncontrolled/untreated *Plasmodium falciparum* infection in humans, is characterized by neurological symptoms, hypothermia, and the sequestration of the infected-erythrocytes (IEs) and platelet microparticles in brain. The sequestration of the IEs in the brain capillaries ensues in hemorrhages, hypoxia, hypoglycemia, convulsions, coma, and ultimately, the death of the patient, if left untreated. Our understanding of human CM (HCM) is rather sparse mainly because of the ethical constraints. So far, knowledge about CM has been obtained from autopsy studies of the brain tissue of CM patients. Therefore, there is urgent need for ideal *in vivo* and/or *in vitro* models of CM, which mimic the HCM, as closely as possible. So far, no ideal experimental CM model has been reported. The critical analysis of the data collected from various *in vivo* and *in vitro* models of CM is expected to augment our understanding of several important aspects of HCM pathogenesis. The identification of biomarkers/biosignatures for the prognosis and diagnosis of HCM are very much warranted. The availability of a suitable experimental model(s) of CM will be helpful in understanding the pathogenesis of HCM, and in the discovery and development of novel therapeutic strategies for HCM.

Keywords: Animal models; Biomarkers; Cerebral Malaria; Hypothermia; *Plasmodium falciparum*; Sequestration

Received: September 26, 2017; **Accepted:** April 19, 2018; **Published:** April 26, 2018

Introduction

The malaria parasite *Plasmodium* has a wide range of host specificity in vertebrates; including primates, rodents, aves and reptiles [1]. These *Plasmodium* parasites are transmitted through mosquito vectors, and have constrained host specificity i.e. the *Plasmodium* species which are confined to rodents only infect the rodents [2,3]. *Plasmodium* infection causes malaria, one of the most prevalent infectious diseases in tropics and sub-tropics [4]. For a long time we knew that naturally, only four *Plasmodium* species (*P. falciparum*, *P. vivax*, *P. malariae* and *P. ovale*) infect humans, and cause malaria [5]. In the recent past; however, three more species of simian malaria parasites (*P. knowlesi*, *P. brasilianum*, and *P. simium*) have been reported to naturally cause malaria in humans [6].

Nevertheless, *P. falciparum* is the only malaria parasite which is highly virulent and often fatal. Cerebral malaria (CM) is a severe neurological complication of malaria in humans and other hosts. During *P. falciparum* malaria, human CM (HCM) is known to

occur which involves the sequestration of IEs in brain capillaries resulting in their blockade [7]. HCM also involves symptoms like hemorrhage, hypoglycemia, hypoxia, convulsions, coma, which eventually lead to the death of the patient if untreated [7]. Several hypotheses have been proposed for the pathogenesis of the HCM, these include (i) mechanical or sequestration hypothesis, (ii) inflammation hypothesis and (iii) hemostasis hypothesis. So far, all the data on HCM has been obtained from the autopsy studies on the brain tissue of CM patients, and, put together, is quite insufficient to draw some meaningful conclusion. Therefore, researchers are still working for an ideal experimental model of CM, which should mimic almost all characteristic features of the HCM. The availability of an experimental model(s) of CM, which closely mimics HCM, will definitely go a long way in augmenting our understanding of the molecular mechanisms of the pathogenesis of HCM, which in turn, is expected to help in the formulation of novel strategies to mitigate and/or to prevent HCM, and also in the discovery and development of novel therapeutic strategies for HCM.

In-vivo/in-vitro Models of CM

Several *in-vivo* and *in-vitro* models are available to study the CM pathogenesis; however, all of them are far from satisfactory. Each of the *in-vivo* or *in-vitro* models has their own strengths and weaknesses. The available *in vivo* CM models include non-human primates, rodents and avian. The integration of the data obtained all these models has been quite helpful in the understanding the pathogenesis of HCM.

In-vivo models

Rodent malaria models: A rodent CM model has been suggested nearly long ago [8]. However, now there are several rodent CM models available to study various aspects of the biology of HCM. A rodent host and malaria parasite combination, based up on its stringency, ultimately determines the outcome of the model i.e. whether it mimic the HCM or not [9]. In murine malarial, the chances of the occurrence of CM incidences and their reproducibility are relatively high. In these models, the immune system is highly active in modulating the pathogenesis and the consequent outcome of CM. The HCM and mouse experimental CM models (ECM) have several similarities, which include (i) the pathology of the nervous system and clinical manifestations of the nervous system malfunction, (ii) disturbed cytokine balance induced brain pathology, (iii) brain endothelial cell involvement in disease pathology and as effector cells in the pathophysiology of malaria, and (iv) similarities in the immune activation pathways [10, 11]. However, every model of murine malaria is not suitable to be developed as a model of ECM; and to act as a model for HCM. It requires very stringent host-parasite combination system, a particular strain of the parasite, the size of the inoculum, the diet of host and the environmental conditions [12-14]. The murine CM was characterized by the inflammation, sequestration of the immune cells in the brain microvessels and vascular leakage of the capillaries due to blood-brain barrier (BBB) infiltration. Thus the murine ECM deviated from the HCM, in terms of an absence of the sequestration of IEs in brain microvessels, little or no BBB dysfunction in the case of the HCM; the vascular leakage of the BBB of murine ECM was similar to the pediatric HCM condition. Despite the vast deviations in the disease pathology/ characteristics and severity between murine ECM and HCM, still, these murine models of ECM are being used to study the CM pathogenesis, to discover the new interventions, and to develop the strategic therapeutic treatments for the HCM. Usually, it is the combination of the rodent malaria parasite and mouse strain which determines the outcome of a particular ECM model viz. i) *P. berghei* ANKA-CBA/ca mice strain represents the neuropathology of the CM characterized by the slight or little IEs sequestration in the microvessels, and cytokines such as TNF- α and IFN- γ are involved in the inflammation and the vascular occlusion of the brain capillaries [15], ii) *P. berghei* ANKA-BALB/c mice strain this combination never been used as model for ECM because BALB/c mice are resistant to develop CM [15], iii) *P.berghei* ANKA-C57BL/6 mice strain also act as a model to study the HCM but the brain vasculature was sequestered with the leucocytes and the cytokines involved in the pathology include TNF- α and IFN- γ [15], iv) *P. berghei* ANKA -DBA/2 mice strain combination represents neuronal pathology of ECM as like as HCM in terms of the recovery

from the CM. The neuronal pathology was mild and recoverable [15], v) *P. berghei* ANKA- BALB/c \times C57BL/6 mice strain will represent the model of age-dependent neuropathology i. e the development of the ECM is based on the age of the mice chosen for the experiment there are both sequestrations of the IEs and there is infiltration of the leucocytes in the microvasculature of the nervous system, IFN- γ was highly expressive in this system [15], vi) *P. berghei* K173 - C57BL/6 mice strain also stand for the model of ECM to investigate the HCM, but the neurological syndrome is due to the sequestration of the activated leucocytes and the IFN- γ involved in the disease pathology [15], vii) the combination of the *P. yoleii* (17XL) - Swiss mice strain stands for the ECM model to access the HCM pathology, neurological sequel is due to the sequestration of the IEs in the microvasculature of the brain [15], viii) the host-parasite system of the *P. yoleii* (17XL) - BALB/c mice strain combination acts as a model of ECM to study the cerebral pathology of the HCM. In this combination system, the IEs were sequestered/cytoadhere to the brain capillary endothelium contributes to pathology [15]. Currently, the *P. berghei* ANKA- C57BL/6 mice model of the ECM considered as a gold standard model to investigate HCM. There are several similarities of characteristics of disease histopathology between *P. berghei* ANKA model ECM and HCM, which are as follows [10,11], i) brain hemorrhage, was observed in both of these two CM conditions as small patches in the brain due to the vascular leakage of the capillaries and venules, ii) cytoadherence/ agglutination/sequestration of the IEs due to their interaction with the microvascular endothelium in both humans and mouse but the extent of the sequestration of the IEs in the mice brain microvessels in case of ECM is limited in nature, iii) occlusion and embolization of the brain microvasculature are common in both HCM and ECM, iv) sequestration of leucocytes in the neuro-capillary network of the brain occurs in both cases of the HCM and ECM but the extent of the leucocytes sequestered more in the case of murine ECM, v) necrosis of the damaged microvessels observed in the both cases of the HCM and ECM, vi) major histocompatibility complexes of class I and II highly expressive and overactive in the cerebral pathology such as CM in both mice and humans, vii) overexpression of the TNF receptors takes place in murine ECM and HCM, viii) and excessive stimulation of the cell adhesion molecules (CAMs) such as ICAM-1, VCAM-1, E-selectin and CD36 occurs predominantly in the HCM; in case of murine ECM; ICAM-1 and VCAM-1 are vastly expressed [15-19].

Avian malaria model(s): Avian *Plasmodium* can be easily transferred by the mosquito vectors whenever there are favorable conditions [20]. *Plasmodium* species are pathogenic to domestic poultry found in Africa, Asia, and South America. Clinical symptoms of avian malaria ranging from no clinical manifestations to severe malaria and death. Infections with the *P. gallinaceum*, *P. juxtranucleare*, and *P. durnae* are most dangerous for the poultry birds because they produce around 90% mortality. *P. gallinaceum* infects chickens and was first described by Brumpt in 1935, formed a foremost model for the systemic study of human malaria [21-23], and governed as a screening model for the studies of the biology of the parasite, immune responses and chemotherapeutic research from 1890 until 1940 [22-24]. This malaria model has also been used in current researches [24-26].

P. gallinaceum infection in chickens represents a satisfactory model for malaria research because of the parasite is in disease pathology and severity such as causing CM [26]. In this study model mortality was proportional to the parasitemia i.e. higher parasitemia correlated to death of infected bird [26]. The body temperature of the bird was positively correlated with parasitemia; increase parasitemia, increase in the body temperature [26]. Hematocrit values were inversely proportional to the parasitemia of the infected bird [26]. The histopathology studies of brain tissue obtained from, the infected bird shown the tissue infiltration of various immune cells and occlusion of the brain microvessels as similar as of the HCM [26].

Non-human primate malaria model(s): A very close genetic and phylogenetic relationship between non-human primate (NHP) hosts and the malarial parasites which infect them, with humans and human malaria parasites makes the NHP malaria models most appropriate and very reproducible models to study various pathogenic, immunological and chemotherapeutic aspects of human malarias. Therefore, it is considered expedient and appropriate to use NHP malaria models as ECM models to mimic human HCM [9]. Though NHPs can be experimentally infected by the malaria parasites despite their natural tendency to get the infection through mosquito bites, the knowledge regarding the disease outcome in these animals remains elusive. Apparently, NHP malaria models seem to be suitable systems for ECM to study HCM; however it is difficult to study the dynamics of CM, especially the onset of the characteristic manifestations. Further, the frequency of occurrence and extent of incidence of CM in NHP malaria models have been observed to be quite diminutive and unpredictable. The acquisition, logistical, maintenance and the related cost considerations involved, coupled with ethical issues regarding their usage for research purposes and the lack of the availability of gene knock-down and knockout models makes NHP malaria models rather unsuitable/un-preferred ECM models [9]. Nevertheless, the NHP host-parasite systems appear suitable for highly effective diagnostic techniques of neuroimaging such as functional MRI etc. to study the disease pathogenesis and to know the degree of severity [27]. There are various host-parasite combinations in NHP malaria models to study the ECM. i) The *P. coatneyi* – *Macaca mullata* (rhesus monkey) shows CM complications such as sequestration of the IEs in the brain microvasculature and expression of cellular adhesion molecules such as ICAM-1, CD36, TSP etc. This model was considered to be an acceptable model to investigate HCM [28-31], ii) *P. fragile* – *Macaca mullata* shows characteristics such as rosetting of IEs and the sequestration of the IEs in the brain microvessels following CAMs are expressed and involved in the pathogenesis like; ICAM-1, CD36; TSP etc. Neurological syndrome characterized by fitting and coma [32-33], iii) *P. falciparum* – *Saimiri sciureus* (squirrel monkey) host – parasite association ECM model characterized by the rosetting of the IE and the sequestration of the IEs occurs in lungs, kidneys and other organs including brain microvessels, but the cell adhesion molecules expression was similar to the other host-parasite systems, expressed CAMs listed as follows ICAM-1, CD36, TSP etc. CM symptoms characterized by fitting and coma [34], iv) *P. knowlesi* – *Macaca mullata* ECM model of NHP malaria model was characterized by the sequestration of the IEs in the

neuronal capillary network and neuronal infection characterized by the mild coma in the infected host animal [35-36], v) *P. coatneyi* – *Macaca fuscata* (Japanese macaques) model of NHP malarial model mimics HCM in terms of rosetting, sequestration, and severity of the neurological syndrome [31], vi) *P. knowlesi* - *Papio anubis* (olive baboon) NHP malaria model has been suggested as an ECM model for HCM, having characteristic sequestration of IEs in brain microvasculature and the neurological symptoms appear during end stage infections [37], vii) *P. falciparum* is known to infect neotropical *Aotus* monkeys (owl monkeys), and causes severe disease complications and death [9].

In vitro models

In vitro models of CM are thought to be very helpful in cell-cell interaction studies *viz.* behavior of IEs, the function(s) of CAMs, the elaboration various of soluble molecules, host cell apoptosis, role of brain parenchymal cells and BBB changes in the CM. Additionally, the *in vitro* models are quite useful in striking a fine balance among the observations of autopsy, animal model and human genetic studies [38-40]. *In vitro* CM models lend additional support to the deep understanding of the different cell-cell interactions, host proteins interactions, and of various variable parasite proteins involved in the cytoadherence. Several *in vitro* models of CM have been reported using different types of cells and cell-lines, which include i) HUVEC (human umbilical vein endothelial cells), ii) HBEC (human brain endothelial cells), iii) HLEC (human lung endothelial cells), iv) HMEC (human mammary endothelial cells), v) stably transfected Chinese hamster ovary (CHO) with CD36 coding genes, vi) stably transfected Chinese hamster ovary (CHO) with ICAM-1 coding genes, vii) monkey brain microvascular endothelial cells (monkey brain MVEC), viii) mouse brain microvascular endothelial cells (mouse brain MVEC), ix) human monocytes, x) human platelets, xi) C32 amelanotic melanoma cells, xii) U937 myelomonocytic cells, xiii) BB19 (immortalized human brain capillary endothelial cell line) and xiv) retinal whole mount method [15].

Mechanisms of CM Pathology

The mechanism(s) CM pathogenesis still remain debatable. However, a few mechanisms have been proposed which include following three hypotheses.

Sequestration hypothesis

In early 1894, in order to explain CM pathogenesis, marchiafava and bignami hypothesized that during *P. falciparum* malaria, IEs sequestered deeply inside the brain microvasculature causing the vascular blockade, hypoxia, hypoglycemia and accumulation of the toxic products such as lactic acid resulting in confusion, convulsions, coma and ultimately death [41,42]. The accumulation of the lactic acid leads to lactic acidosis in the CM; due to hypoperfusion of the particular brain tissue, tissue generates its energy needs through anaerobic respiration. Though this contention appeared to support sequestration hypothesis, the exact mechanism of hyperlactatemia in CM appear to be quite complicated and need not to be linked to anaerobic respiration [43]. Elevated lactate levels seem to play a significant role in CM pathogenesis [44] can be correlated with;

i) decreased oxygen supply, ii) alterations in the redox status, iii) convulsions and iv) may be due to decreased clearance by the liver [45]. *P. falciparum* infection causes structural and functional alterations in both uninfected and infected erythrocytes, hindering their movement in brain capillaries of 3-7 micron diameter; the rigid RBCs of diameter 7.5 microns will occlude the capillaries and resulting in the mechanical obstruction of the brain capillaries [46]. The integration of the parasitic proteins in the RBC membrane results in the increased rigidity of the membrane of IEs [47]. The uninfected erythrocyte rigidity is due to the excessive oxidative stress [46]. In ECM, there appears to be an inverse relationship between capillary occlusion and the concentration of the functional capillaries [48]. The sequestration hypothesis thus appears to be based on the proportionality of levels of parasitemia and the degree of obstruction of the brain microvasculature. However, on the contrary, no such clinical relationship between the higher parasitemia and the mortality has been reported. It should be noted here that to observe sequestration is not a practical possibility in postmortem studies of each and every *P. falciparum* malaria patient [48,49].

Inflammation hypothesis

In 1948, Brian Maegraith proposed the involvement of inflammatory mediators in systemic inflammatory reaction to malaria pathogenesis leading to failure of vital organs and death [50]. The malaria toxins, which mainly contain glycoposphoinositol, are involved in causing disturbance in the balance of the pro-inflammatory/anti-inflammatory mediators. The increase in oxidative damage is due to the excessive production of the superoxide and nitric oxide (NO) molecules [51]. The so generated pro-inflammatory cytokines such as TNF- α involved in the enhancement of the turnover of the endothelial cell adhesion molecules (eCAMs) such as ICAM-1, one of the key molecules involved in CM pathogenesis. The regulating cytokines for the production of the eCAMs are specific to particular adhesion molecule and to a particular tissue [19]. Experimental proofs have been generated by using the knockout models which suggest the association of immune activated inflammatory cascade and the progression of the CM pathogenesis [52]. The presence of high levels of inflammatory mediators in *P. vivax* malaria indicates that though the participation of the inflammatory cytokines required, but not enough for CM pathogenesis. The outcomes from the clinical trials involving the anti-inflammatory therapy including the antibody (anti-TNF- α monoclonal antibody), pentoxifylline (synthesis inhibitor) and corticosteroid (dexamethasone) have shown that anti-inflammatory therapy seem to be unable to provide the protection from the disease pathology. Moreover, there are some irrational observations during clinical trials [53,54]. The high NO production is also thought to play a role in the CM pathogenesis, which in turn supports, the inflammatory hypothesis. The pleiotropic functions of NO are also being considered to be responsible for pathogenesis of CM [55-61]. The heme released during rupture of the IEs reacts with NO and scavenges it, and thus makes it less available for biological functions [62,63]. Free NO is quenched by superoxide ion due to rigorous oxidative stress in the malignant *P. falciparum* infections [64-66].

Hemostasis hypothesis

The presence of small patches of hemorrhages in the brain, retinal hemorrhages, and vascular leaks are signs of the hemostatic dysfunction in CM. *P. falciparum* malaria patient's abnormalities such as prolonged bleeding, prolonged prothrombin, partial prothrombin times and presence of coagulopathy have been observed, and point out the faults in the factors V, VII, and IX required for the coagulation [67,68]. The low concentrations of anticoagulant proteins and C-reactive protein have been observed in *P. falciparum* malaria [69,70]. Platelets are chief effector cells of hemostasis system and associated with pathogenesis of CM. Immune activated platelets release chemokines, cytokines and other immune modulator molecules from their cytoplasmic granules [71]. Thrombocytopenia and its extent have always been linked to the pathogenesis of the CM [72-74]. Certainly, thrombocytopenia enhances the probability of the bleeding. The platelet adhesion to the endothelium of the brain microvasculature contributes to the occurrence of CM, similarly to the sequestration of the IEs. Further, platelet microparticles may also mediate between endothelium and leukocytes [75]. Microparticles of EC origin are found extensively in the patients with *P. falciparum* infection, can be linked with the progression of the pathology [76,77]. It is important to note here that no single hypothesis can explain the mechanisms of the CM pathogenesis; apparently, at best, these proposed hypothetical mechanisms seem to be dependent on each other and are not exclusive. Extensive studies to unravel the underlying mechanisms are very much warranted.

Factors Influencing the Outcome of CM

A large number of factors can be surmised which influence, directly and/or indirectly, the onset, severity, and outcome in ECM models and HCM. These factors may influence CM both to detriment (aggravate the disease) or benefit (resolve the disease) of the host. These factors may helpful for the parasite in its efforts to evade the mounting immunological onslaught of the host.

Cell adhesion molecules (CAMs)

Cell adhesion molecules are major factors which influence the sequestration of IEs, adhesion of the immune cells and platelets to the endothelium of the brain capillaries. The elevated CAM levels are thought to be augment the communication between the attracted immune cells and endothelium, and are also considered to be instrumental in plugging the leakage in cerebral capillaries [19,78]. ICAM-1, CD36 are prominent among the CAMs act as ligands for the PfEMP-1 protein on the surface of *P. falciparum* IEs; mediates the sequestration and adhesion of the IEs to the endothelium of the brain capillaries. Increased IEs sequestration ensues in vascular occlusion, reduced flow of blood, hypoxia, and some other characteristic features (acidosis, hypoxia, and ischemia) of HCM [19,78,79]. Sequestration thus causes damage to the host endothelium, and leads to the apoptosis of the EC of brain microvasculature which may results in vascular leakage, ultimately the dysfunction of the BBB [19]. The elevated CAM level-induced amplification of sequestration may augment elaboration of inflammatory cytokines, and chemokines from

the EC, which, in turn, may trigger uncontrolled inflammatory cascade causing additional damage to the BBB [80]. Significantly increased levels of ICAM-1 expression, and TNF- α and IL- β levels especially in the cerebellum, have been observed in the brain tissues of children who died of CM [81]. Further, the role of ICAM-1 has been documented in the development of the CM. ICAM-1 deficient mice display resistance to the CM, showed diminished levels of serum TNF- α , lack of immune cell sequestration in brain microvasculature, and no or reduced damage to the BBB when compared to their wild-type littermates [82]. These observations suggest that the CAMs are detrimental in CM.

Cytokines

Cytokines are low molecular weight glycoprotein molecules, regulators of the host response to infection, inflammation and immune response and several other processes. Whereas the induction and involvement of pro-inflammatory cytokines ensues in pathological conditions which are detrimental to host, anti-inflammatory cytokines involved in the reduction of inflammation and augmentation of healing process. Thus, during infection and inflammatory conditions, anti-inflammatory cytokines play protective role(s), pro-inflammatory cytokines aggravate both infection and disease progression. Nevertheless, in *P. falciparum* malaria patients, both pro- and anti-inflammatory cytokines play their specific role(s) both in the onset and progression of CM, and also influence the end result of CM [83].

TNF- α

Tumor necrosis factor- α (TNF- α , catechin) is an important pro-inflammatory Th₁ cytokine which is involved in several biological functions and is extensively involved inflammation. There are several lines of evidence which support the role of TNF- α in the pathogenesis of the CM [83]. Elevated levels of TNF- α was recorded in several cases of CM, and administration of the anti-TNF- α antibody has been reported to mitigate neuronal symptoms and pathology [83]. Elevated levels of soluble forms of TNFR₁ and TNFR₂ are found in the plasma samples of both adult and paediatric patients [84], and also in the case of murine CM [85]. TNFR₂ has proven the potential to cause CM pathology because the TNFR₂ gene knockout and not the TNFR₁ knockout mice were protected from the CM pathology [85]. The elevated expression of the TNFR₂ has been reported in the brain endothelium of the Malawian children with fatal malaria [86].

IFN- γ

The interferons (IFNs) were discovered because of their property to interference with the viral replication [87]. IFNs were categorised into two groups i) type I and ii) type II based on their receptor pharmacology, specificity and sequence homology. The type I interferons includes IFN- α , IFN- β , IFN- ω and IFN- τ . IFN- γ is the only type II interferon and it phenotypically dissimilar to the type I interferons [88]. IFN- γ acts in autocrine or paracrine manner, locally [89]. IFN- γ has immunomodulatory functions, and its production is regulated by the cytokines IL-12 and IL-18 [90]. In murine ECM, IFN- γ has been observed to be related with mortality and disease pathology, and antibodies against the IFN- γ are known to protect the mice from mortality and

the CM development. Mice knockout for both IFN- γ and IFN- γ R have been found defiant to develop CM [79,91]. Acute malaria patients from both South East Asia [92] and African regions [93] have been reported to have high IFN- γ plasma levels. The Gambians, heterozygous for the IFN receptor polymorphism, show low probability of developing CM and its related mortality [94]. Thus observations from both human and murine malaria studies strongly suggest the role of IFN- γ in the CM pathogenesis.

IL-6

IL-6 is an anti-inflammatory cytokine involved in host immune responses, acute inflammatory reactions and hematopoiesis [95]. Significant increase in the serum IL-6 levels has been observed *P. berghei* ANKA infected mice, with or without CM involvement. Severe *P. falciparum* malaria patients also show elevated IL-6 levels [96]. IL-6 has been reported to be involved in the stimulation of the polyclonal B cells, in severe malaria patients [97]. Contrastingly, there are evidences which suggest that IL-6 has nothing to do with the pathogenesis of murine CM [98] There is no deviation of the expression of IL-6 mRNA levels in both CM vulnerable and CM defiant infections of *P. berghei* ANKA [99]. The administration of IL-6 neutralizing antibodies during murine CM did not protect the mice from the disease symptoms and associated mortality, which demonstrates that IL-6 is not involved in CM [98].

IL-10

IL-10 is known to downregulate the production of TNF- α and IL-1 [100]. In malaria, IL-10 has been shown to have some protective roles. The exogeneous administration of IL-10 has been shown to protect against the development of CM in susceptible *P. berghei* ANKA infected mice; however, paradoxically, in mice resistant to *P. berghei* ANKA infection, administration of neutralising antibody lead to development of CM pathology [101]. Human malaria studies indicate the host protective activity of the IL-10, as shown by the low levels of IL-10 in CM patients as compared to that in uncomplicated malaria [102]. Further, IL-10 seems to protect against CM pathology by restraining the production of TNF- α and GM-CSF, both of which considered responsible for the CM complications [83,103]. Additionally, IL-10 exerts protection against CM by blocking the production of the several chemokines [104], which attract monocytes and CD8⁺ T cells responsible for the blockade of cerebral microvasculature during CM [79, 105-107]. IL-10 thus appears to be a host protective cytokine during the CM pathogenesis.

IL-1

IL-1 is a pro-inflammatory cytokine, a principal mediator in the acute inflammatory response, and potential mediator of tissue malfunction and devastation. IL-1 is responsible for mediating neuroinflammation and neurodegeneration [108]. IL-1 plays a highly critical role in the production of the Th17 cells from the T cell population [109]. The role of IL-1 in the CM pathogenesis is quite complicated. The administration of the low doses of IL-1 offered protection against lethal CM [110]. Treatment with recombinant IL-1 receptor antagonist inhibited the progression of infection, and averted the development of the CM complications [97].

IL-4

IL-4 is involved in I_g isotype switching, expression of major histocompatibility complex class II by B cells and the differentiation of T cell subsets [111]. Th₂ cell activation seems important for the clearance of the late stage malaria parasites, and IL-4 activation thus appears to protect against malaria, but even IL-4 scarce mice are able to clear the primary infections of *P. chabaudi* [112]. Addition of recombinant human IL-4 to the *in vitro* culture of the *P. falciparum* inhibited macrophage killing of parasite, thus helped the parasite to escape the immune system [113]. There is decreased expression of IL-4 mRNA in the brains of the mice with CM, which indicates the protective role of IL-4 during CM pathology [99]. Nevertheless, uncertainty continues regarding the role of the IL-4 in the pathogenesis, and regulation of the outcome of the CM.

Vasomodulatory agents

The pathophysiological changes of the CM are associated with the brain microvasculature, thus the recent research regarding the CM primarily focused on the endogenous Vasomodulatory agents such as endothelin-1 and NO.

Endothelin-1

Endothelin-1 (ET-1) is a potent vasoconstrictor secreted by the endothelial cells of the vasculature. The increased ET-1 levels have been observed in the brains of *P. berghei* ANKA-infected mice [114], which are associated with cerebral hypoperfusion and immunopathology of CM [114, 115]. The vasculopathy of CM is mediated by the ET-1, through its action on the endothelin receptor type A (ETA) [116]. The ETA antagonism decreased the haemorrhage and vascular occlusion during CM, and enhanced the survival rate of mice given artemisinin supportive therapy [116]. The IE sequestration-induced endothelium over stress causes noticeable raise in ET-1 levels, and up-regulates the CAM expression [114]. The CAM up-regulation triggers the cerebral sequestration of the IEs, monocytes and platelets and other immune cells, which results in the vascular occlusion, cerebral hypoperfusion, hypoxia, ultimately leads to ischemia and death [117]. Thus it can be concluded that the hyper-expression of the ET-1 is detrimental in CM.

Nitric oxide

The activities of nitric oxide (NO) include immunomodulatory, vasomodulatory, signal transduction and cell growth regulatory functions [118]. The role of NO in CM is matter of conjecture and incongruity, as both up- and down-regulation in NO levels cause damaging effect of CM. Increased NO levels result in higher intracranial pressure, interference in the neuronal signal transmission and weight loss [119]. On the other hand, decreased NO levels results in endothelial damage and dysfunction. Decrease in NO levels during severe malaria includes hypoarginemia and NO quenching [120,122]. Decreased NO levels lead to CM, haemorrhages, and vessel collapse [16,80,114,120]. Curiously, in both HCM and murine CM, arginine treatment restored the NO levels, reorganised endothelial function, and thus normalized BBB functions [120-124]. In *P. gallinaceum*-infected chickens,

treatment with amino guanidine, an NO synthesis inhibitor, increased resistance to infection due to reduced inflammation, anaemia and thrombocytopenia [125].

Prostaglandins

Prostaglandins (PG) are generated by the action of cyclic oxygenases (COX) on the arachidonic acid (AA), and are involved in macrophage stimulation, vascular integration, erythropoiesis, and fever, and pro-inflammatory responses to the infection [126]. The children having malaria-induced severe malaria have concealed bicyclo PGE₂ plasma levels and repressed leukocyte COX-2 gene [126]. Suppressed bicyclo PGE₂ levels lead to severe malaria and CM [126]. Aspirin treatment of ICR mice during ECM aggravated the mortality rate due to inhibition of the PG synthesis [127]. These observations indicate a protective role of PG in CM.

Leukotrienes

Leukotrienes (LT) have an important role in innate and adaptive immune system responses, and are implicated in several inflammatory and pathological conditions, and infectious diseases [128,129]. Cysteine Leukotrienes are responsible for the vascular permeability and edema and the expression of the adhesion molecules and NO production [128]. In *P. berghei* ANKA-induced murine ECM, elevated serum LTB₄ levels have been observed, and aspirin treatment drives the AA metabolism towards the 5-LOX enzyme [130], leading to rapid rise in parasitemia and aggravated mortality during CM [127]. LTB₄ is a potent inducer of Th₁ cytokines such as IFN- γ [131] overproduction of this Th1 cytokine-associated with the severity of CM pathogenesis. LT may thus have a deleterious role(s) in the CM.

Glycophosphatidylinositol

Glycophosphatidylinositol (GPI), a ubiquitous molecule, is present in outer cell membrane of all eukaryotes. Studies related to *In vivo* and *in-vitro* studies suggest a potential role(s) of *Plasmodium* GPIs in pathogenesis and severity of the disease. Interaction of *P. berghei* and *P. yoelli* IEs with macrophages, *in vitro*, ensued in the elaboration of TNF- α [132], and similar results were obtained with human monocytes with the *P. falciparum* IEs [133]. Purified GPIs, following injection in thioglycolate-primed mice, caused acute malaria manifestations such as fever and hypoglycemia [134], similar results were observed in unprimed mice [135]. Neutralisation of the *Plasmodium* GPIs with monoclonal antibodies nullified the TNF- α production induced by the whole parasite extracts *in vitro* [136]. *P. falciparum* GPIs cause the elevation of the endothelial cell expression of the ICAM-1, VCAM-1 and E-selectin via TNF- α and IL-1[137]. GPIs of *Plasmodium* appear to cause pathogenesis of CM.

Hemozoin

Hemozoin (Hz), the malaria pigment, is a metabolic waste product produced due to digestive catabolism of the haemoglobin in IEs by parasite [138]. The Hz taken-up by phagocytic cells via phagocytosis of the IEs or free Hz released after schizont rupture [139]. Hz thus released alters the cellular metabolism through generation of free radical lipid peroxides from the AA

[140]. Phagocytic up-take of the Hz by circulating neutrophils and macrophages has been linked to the severity of the malaria [141,142]. The Hz formation has also been linked with severe malarial anemia [143], immunosuppression [139], and cytokine dysregulation [144]. Cytokine dysregulation leads to the alterations in the balance of the pro/anti-inflammatory cytokines, which results in aggravating CM during *P. falciparum* malaria.

Alterations in signalling and biological pathways

In severe malaria, parasite-induced disturbances in the host homeostasis system lead to the alterations in the signalling and biological pathways, which in turn, badly influence the pathophysiology of the CM.

Kynurenine pathway of tryptophan metabolism

Currently, there appears to be sufficient information which supports the view that during HCM and Murine CM, the kynurenine pathway of tryptophan metabolism is stimulated. The Indoleamine oxygenase (IDO), rate-limiting enzyme of the pathway, has been reported to be up-regulated in the brains of mice suffering from CM [145]. Excessive stimulation of the enzyme IDO alters the ratio of the neuroprotective metabolite kynurenic acid (KA) and neuro-excitotoxic metabolite quinolinic acid (QA), and accumulation of QA is known to occur during CM, which leads to neuronal damage [145]. The heightened QA levels have been observed in the cerebrospinal fluid of adult human CM [146], and paediatric CM patients [147]. Murine CM studies, therefore, suggest that inhibition of the Kynurenine pathway extended the survival of the treatment mice three-times longer, as compared to the untreated ones [148].

Angiotensin-Tie 2 signalling pathway

The CM pathology, mainly involves the structural and functional alterations in cerebral microvasculature, both of which can be maintained by factors such as NO, ET-1 etc. The angiotensin-Tie 2 signalling pathway is one of the regulators of the microvasculature of the brain. Basically angiotensin-1 (Ang-1) interacts with EC tie 2, and controls the stimulation of the endothelium and maintains the survival of the ECs [18]. Paradoxically, angiotensin-2 (Ang-2) functions just the other way round of Ang-1[18]. Ang-1/Ang-2 ratio change towards Ang-2 causes sensitization of the endothelium, which results in increased generation of the CAMs, and thus, further aggravates the CM complications [149,150].

Rho-kinase signalling pathway

The BBB integrity is maintained by tight junctions and adherent junctions, which are associated with small GTPase protein rho. Stimulated rho A/rho kinase pathway results in the endothelial dysfunction [151]. Fasudil, a rho-kinase inhibitor, reduced the EC apoptosis, NF- κ B activation [152], and prolonged the survival, and can thwart the development of the CM in murine ECM [153].

Toll-like receptor signalling pathway

Toll-like receptors (TLRs) play major role(s) in the generation of primary innate immune responses against the pathogens. TLRs stimulation also activates the production of IFNs, and secretion of Th1 and Th2 cytokines [154]. Most of the TLRs intracellular signalling is MyD88 adaptor molecule dependent [155]. Pro-inflammatory responses to the malaria parasites mediated through the TLR4, TLR9 and adaptor molecule MyD88, after interaction between antigen presenting cell-dendritic cell and *P. berghei*/*P. chabaudi* IEs [156]. TLR2 and TLR9 receptors are involved in the pathogenesis of CM, they aggravate CM related mortality [155].

Heme/Heme oxygenase-1 signalling pathway

Heme oxygenase (HO-1) degrades free heme, which ensues in the formation of bile pigment, biliverdin, free iron, and carbon monoxide (CO) [157]. The heightened HO-1 levels during oxidative stress, exposure to UV rays, heavy metal toxicity and inflammatory pathological conditions, contribute to defence against deleterious effects of these factors. Excessive free heme due to severe hemolysis of IEs and uninfected RBCs during malaria, leads to inflammation and contributes cerebral pathology [158-160]. Recent ECM studies *P. berghei* ANKA show that upregulation of HO-1 enzyme levels and its activity protected the host from the developing ECM. Crossing out of HO-1 and inhibition of its activity augmented cerebral pathology by 83% and 78% respectively [159]. Taken together, these observations, in no uncertain terms, suggest the protective role of enzyme HO-1 during CM.

Biomarkers of CM

So far, there are no explicit biomarkers for the diagnosis of the HCM or ECM, but some recent studies have recognised some serological factors to have the potential to function as biomarkers. It is believed that identification and application of potential practical biomarkers/biosignatures will go a long way in the diagnosis and prognosis and for the therapeutic assessment of a drug or vaccine for CM. In order to identify the biomarkers for the particular disease, it is necessary to have a deep knowledge of disease pathogenesis and of pathophysiological changes during the progression of the disease. However, the pathogenesis of the CM still remains to be properly understood and expounded. Some recent studies in Ghana [161] and India [162] have exposed the remarkable relationship between chemokine interferon-inducible protein-10 (CXCL10) and CM severity, which proposes the potential of the CXCL10 to be used as a practical biomarker for the assessment or quantification of the severity of the CM. Further, there are several serological factors whose concentrations vary, which can potentially be used to discriminate CM for non-CM conditions. These serological factors include elevated plasma/serum levels of the CXCL10, sFas, sTNF-R2 [161, 162], IL-8, IL-1ra [163], and diminished levels of RANTES [164] and vascular endothelial growth factor (VEGF) in CM patients. Till date, only the endothelial regulators Ang-1 and Ang-2 [150,165], and chemokine CXCL10 [166] have been evaluated for their potential to diagnose CM. Erythropoietin

circulating levels also have been thought of as a potential prognostic biomarker for CM; higher levels are associated with 70% less chances of developing neurological sequelae, whereas low levels indicate risk of neurological involvement [167-169]. Increased CXCL10 levels in CSF and peripheral serum samples of CM patients who have died, compared to that in the survivors, suggests its potential to be a prognostic biomarker for CM. Thus all these factors can potentially be developed as diagnostic biomarkers/ biosignatures for CM.

References

- 1 Hayakawa T, Culleton R, Otani H, et al. (2008) Big bang in the evolution of extant malaria parasites. *Mol Biol Evol* 25: 2233-2239.
- 2 Levine ND (1988) The protozoan phylum Apicomplexa. Boca Raton.
- 3 Coatney GR, Collins WE, Warren M, et al. (2003) CDROM. The primate malarias. Atlanta.
- 4 Centers for Disease Control and Prevention.
- 5 World Health Organization
- 6 Cox-Singh J, Singh B (2008) Knowlesi malaria: newly emergent and of public health importance? *Trends Parasitol* 24: 406-410.
- 7 Newton CR, Hien TT, White N (2000) Cerebral malaria. *J Neurol Neurosurg Psychiatry* 69: 433-441.
- 8 Yoeli M, Hargreaves BJ (1974) Brain capillary blockage produced by a virulent strain of rodent malaria. *Science* 184: 572-573.
- 9 Craig AG, Grau GE, Janse C, et al. (2012) The role of animal models for research on severe malaria. *PLoS Pathog* 8.
- 10 Hunt NH, Grau GE (2003) Cytokines: accelerators and brakes in the pathogenesis of cerebral malaria. *Trends Immunol* 24: 491-499.
- 11 Taylor-Robinson, AW (2010) Validity of modelling CM in mice: argument and counter argument. *J Neuroparasitology* 1: 1-5.
- 12 Amani BMI, Pied S, Marussig M, et al. (1998) Cloned lines of *Plasmodium berghei* ANKA differ in their abilities to induce experimental cerebral malaria. *Infect Immun* 66: 4093-4099.
- 13 Gilks CF, Jarra W, Harvey WK, et al. (1989) Host diet in experimental rodent malaria: a variable which can compromise experimental design and interpretation. *Parasitology* 2:175-177.
- 14 Herbas MS, Okazaki M, Terao E, et al. (2010) Alpha-Tocopherol transfer protein inhibition is effective in the prevention of CM in mice. *A J Clin Nutrition* 91: 200-207.
- 15 Ozwara H (2007) Experimental models of cerebral malaria. Malaria programme, Institute of Primate Research, Nairobi.
- 16 Zanini GM, Cabrales P, Barkho W, et al. (2011) Exogenous nitric oxide decreases brain vascular inflammation, leakage and venular resistance during *Plasmodium berghei* ANKA infection in mice. *J Neuroinflammation* 8: 66.
- 17 Grau GE, Pigué PF, Engers HD, et al. (1986) L3T4+ T lymphocytes play a major role in the pathogenesis of murine cerebral malaria. *J Immunol* 137: 2348-2354.
- 18 Conroy AL, Glover SJ, Hawkes M, et al. (2012) Angiopoietin-2 levels are associated with retinopathy and predict mortality in Malawian children with cerebral malaria: a retrospective case-control study. *Crit Care Med* 40: 952-959.
- 19 Bauer PR, Specian RD, Granger DN, et al. (2002) Regulation of endothelial cell adhesion molecule expression in an experimental model of cerebral malaria. *Microcirculation* 9: 463-470.
- 20 Redig PT (1993) Avian malaria. *Proceedings of Association of Avian Veterinarians* 1: 173-181.
- 21 Paraense WL (1946) Ações patogênicas das formas exo-eritrocitárias do *Plasmodium gallinaceum*. *Memorial Institute of Oswaldo Cruz* 44: 147-91.
- 22 Garnham PC (1966) Malaria Parasites and other Haemosporidia. Oxford, Blackwell Scientific Publications.
- 23 Williams RB (2005) The efficacy of a mixture of trimethoprim and sulphadoxine against *P. gallinaceum* malaria in the domesticated fowl *Gallus gallus*. *V Parasitology* 129: 193-197.
- 24 Slater LB (2005) Malarial birds: modeling infectious human disease in animals. *Bull Hist Med* 79: 261-294.
- 25 Braga EM, Silveira P, Belo NO, et al. (2011) Recent advances in the study of avian malaria: an overview with an emphasis on the distribution of *Plasmodium* spp. in Brazil. *Mem Inst Oswaldo Cruz* 1: 3-11.
- 26 Macchi BM, Quaresma JA, Herculano AM, et al. (2010) Pathogenic action of *Plasmodium gallinaceum* in chickens: brain histology and nitric oxide production by blood monocyte-derived macrophages. *Veterinary Parasitology* 172: 16-22.
- 27 Kawai S, Sugiyama M (2010) Imaging analysis of the brain in a primate model of cerebral malaria. *Acta Trop* 114: 152-156.
- 28 Udonsangpetch R, Brown AE, Smith CD, Webster (1991) Rosette formation by *Plasmodium coatneyi*-infected red blood cells. *Am J Trop Med Hyg* 44: 399-401.
- 29 Aikawa M, Brown A, Smith CD, et al. (1992) A primate model for human CM: *Plasmodium coatneyi* infected rhesus monkeys. *Am J Trop Med Hyg* 46: 391-397.
- 30 Nakano Y, Fujioka H, Luc KD, et al. (1996) A correlation of the sequestration rate of *Plasmodium coatneyi*-infected erythrocytes in cerebral and subcutaneous tissues of a rhesus monkey. *Am J Trop Med Hyg* 55: 311-314.
- 31 Kawai S, Kano S, Suzuki M (1995) Rosette formation by *Plasmodium coatneyi*-infected erythrocytes of the Japanese macaque (*Macaca fuscata*). *Am J Trop Med Hyg* 53: 295-299.
- 32 Fremont HN, Miller LH (1975) Deep vascular schizogony in *Plasmodium fragile*: organ distribution and ultrastructure of erythrocytes adherent to vascular endothelium. *Am J Trop Med Hyg* 24: 1-8.
- 33 Fujioka H, Millet P, Maeno Y, Nakazawa S, Ito Y (1994) A nonhuman primate model for human CM: rhesus monkeys experimentally infected with *Plasmodium fragile*. *Exp Parasitology* 78: 371-376.

Acknowledgment

We are grateful to Prof. Raghu Ramarao Akkinepalli, Director, National Institute of Pharmaceutical Education and Research (NIPER), for his help and encouragement. Mr. Bhanu Prakash is grateful to the NIPER for the award of a Senior Research Fellowship. This is NIPER communication No. 499.

- 34 Gysin J (1992) Mechanisms of protective immunity against asexual blood stages of *Plasmodium falciparum* in the experimental host Saimiri. *Memorial Institute of Oswaldo Cruz* 87: 407-412.
- 35 Al-Khedery B, Barnwell JW, Galinski MR (1999) Antigenic variation in malaria: a 39 genomic alteration associated with the expression of a P. knowlesi variant antigen. *Mol Cell* 3: 131-141.
- 36 Korir CC, Galinski MR (2006) Proteomic studies of P. knowlesi SICA variant antigens demonstrate their relationship with P. falciparum EMP1. *Infect Genet Evol* 6:75-79.
- 37 Ozwara H, Langermans JA, Maamun J, Farah IO (2003) Experimental infection of the olive baboon (*Papio anubis*) with P. knowlesi: severe disease accompanied by cerebral involvement. *Am J Trop Med Hyg* 69: 188-194.
- 38 Udeinya IJ, Schmidt JA, Aikawa M, et al. (1981) Falciparum malaria-infected erythrocytes specifically bind to cultured human endothelial cells. *Science* 213: 555-557.
- 39 Berendt AR, Simmons DL, Tansey J, Newbold CI, Marsh K (1989) Intercellular adhesion molecule-1 is an endothelial cell adhesion receptor for *Plasmodium falciparum*. *Nature* 341: 57-59.
- 40 Oquendo P, Hundt E, Lawler J, Seed B (1989) CD36 directly mediates cytoadherence of *Plasmodium falciparum* parasitized erythrocytes *Cell* 58: 95-101.
- 41 Idro R, Jenkins NE, Newton CR (2005) Pathogenesis, clinical features, and neurological outcome of cerebral malaria. *Lancet Neurol* 4: 827-840.
- 42 Miller LH, Baruch DI, Marsh K, et al. (2002) The pathogenic basis of malaria. *Nature* 415: 673-679.
- 43 White NJ, Ho M (1992) The pathophysiology of malaria. *Adv Parasitol* 31: 83-173.
- 44 Marsh K, Forster D, Waruiru C, et al. (1995) Indicators of life-threatening malaria in African children. *N Eng J Med* 332: 1399-1404.
- 45 Maitland K, Newton CR (2005) Acidosis of severe falciparum malaria: heading for a shock? *Trends Parasitol* 21: 11-16.
- 46 Dondorp AM, Kager PA, Vreeken J, White NJ (2000) Abnormal blood flow and red blood cell deformability in severe malaria. *Parasitol Today* 16: 228-232.
- 47 Glenister FK, Coppel RL, Cowman AF, et al. (2002) Contribution of parasite proteins to altered mechanical properties of malaria-infected red blood cells. *Blood* 99:1060-1063.
- 48 Clark IA, Awburn MM, Whitten RO, et al. (2003) Tissue distribution of migration inhibitory factor and inducible nitric oxide synthase in falciparum malaria and sepsis in African children. *Malaria J* 2: 6.
- 49 Taylor TE, Fu WJ, Carr RA, Whitten RO, Mueller JS, Fosiko G (2004) Differentiating the pathologies of CM by postmortem parasite counts. *Nature Med* 10: 143-145.
- 50 Maegraith B (1948) Pathological processes in malaria and blackwater fever. *Ind Med Gaz* 84: 221.
- 51 Nebl T, De Veer, Schofield L (2005) Stimulation of innate immune responses by malarial glycosylphosphatidylinositol via pattern recognition receptors. *Parasitol Today* 130.
- 52 Schofield L, Grau GE (2005) Immunological processes in malaria pathogenesis. *N Rev Im* 5: 72-35.
- 53 Van Hensbroek MB, Palmer A, Onyiorah E, Schneider G (1996) The effect of a monoclonal antibody to tumor necrosis factor on survival from childhood CM. *J Infect Dis* 174: 1091-1097.
- 54 Prasad K, Garner P (2000) Steroids for treating cerebral malaria. *Cochrane Database Syst* 2: CD000972.
- 55 Anstey NM, Weinberg JB, Hassanali MY, et al. (1996) Nitric oxide in Tanzanian children with malaria: inverse relationship between malaria severity and nitric oxide production/nitric oxide synthase type 2 expression. *J Exp Med* 184: 557-567.
- 56 Bogdan C (2001) Nitric oxide and the immune response. *Nat Immunol* 2: 907-916.
- 57 Tuteja N, Chandra M, Tuteja R, et al. (2004) Nitric oxide as a unique bioactive signalling messenger in physiology and pathophysiology. *J Bio Biotech* 4: 227-237.
- 58 Laroux FS, Lefer DJ, Kawachi S, et al. (2000) Role of nitric oxide in the regulation of acute and chronic inflammation. *Anti Re Signal* 2: 391-396.
- 59 Loscalzo J (2001) Nitric oxide insufficiency, platelet activation, and arterial thrombosis. *Cir Res* 8: 756-762.
- 60 van der Veen RC (2001) Nitric oxide and T helper cell immunity. *Int Immunopharmacol* 1: 1491-1500.
- 61 Lopansri BK, Anstey NM, Weinberg JB, et al. (2003) Low plasma arginine concentrations in children with CM and decreased nitric oxide production. *Lancet* 361: 676-678.
- 62 Rother RP, Bell L, Hillmen P, et al. (2005) The clinical sequelae of intravascular hemolysis and extracellular plasma haemoglobin: a novel mechanism of human disease. *JAMA* 293: 1653-1662.
- 63 Reiter CD, Wang X, Tanus-Santos JE, et al. (2002) Cell-free haemoglobin limits nitric oxide bioavailability in sickle-cell disease. *Nature Med* 8: 1383-1389.
- 64 Estévez AG, Jordán J (2002) Nitric oxide and superoxide, a deadly cocktail. *Sci* 962: 207-211.
- 65 Rubanyi GM, Vanhoutte PM (1986) Superoxide anions and hyperoxia inactivate endothelium-derived relaxing factor. *Am J Physiology* 250: 822-827.
- 66 Thomas SR, Chen K, Keaney JF (2003) Oxidative stress and endothelial nitric oxide bioactivity. *Anti Re Signaling* 5: 181-194.
- 67 Rojanasthien S, Surakamolleart V, Boonpucknavig S, et al. (1992) Haematological and coagulation studies in malaria. *J Med A Thailand* 75: 190-194.
- 68 Srichaikul T, Pulket C, Sirisatepisarn T, et al. (1988) Platelet dysfunction in malaria. *Southeast Asian J Trop Med Public Health* 19: 225-233.
- 69 Essien EM (1989) The circulating platelet in acute malaria infection. *Br J Haematol* 72: 589-590.
- 70 Essien EM, Ebhota MI (1983) Platelet secretory activities in acute malaria (*Plasmodium falciparum*) infection. *Acta Haematologica* 70: 183-188.
- 71 Weyrich AS, Zimmerman GA (2004) Platelets: signaling cells in the immune continuum. *Trends Immunol* 25: 489-495.
- 72 Chinowsky MS (1991) Cerebral falciparum malaria mimicking thrombotic thrombocytopenic purpura. *S Med J* 84: 374-378.
- 73 Horstmann RD, Dietrich M, Bienzle U, Rasche H (1981) Malaria-induced thrombocytopenia. *Blut* 42: 157-164.
- 74 Horstmann RD, Dietrich M (1985) Haemostatic alterations in malaria correlate to parasitaemia. *Blut* 51: 329-335.
- 75 Huber J, Vales A, Mitulovic G, et al. (2002) Oxidised membrane vesicles and blebs from apoptotic cells contain biologically active oxidised

- phospholipids that induce monocyte–endothelial interactions. *Arte Throm Vas Bio* 22: 101-107.
- 76 Combes V, Taylor TE, Juhan-Vague I, et al. (2004) Circulating endothelial microparticles in Malawian children with severe falciparum malaria complicated with coma. *JAMA* 291: 2542-2544.
- 77 Combes V, Coltel N, Alibert M, et al. (2005) ABCA1 gene deletion protects against CM: potential pathogenic role of microparticles in neuropathology. *Am J Pathology* 166: 295-302.
- 78 Beare NA, Harding SP, Taylor TE (2009) Perfusion abnormalities in children with CM and malarial retinopathy. *J Infect Dis* 199: 263-271.
- 79 Yañez DM, Manning DD, Cooley AJ, et al. (1996) Participation of lymphocyte subpopulations in the pathogenesis of experimental murine cerebral malaria. *J Immunol* 157: 1620-1624.
- 80 Cabrales P, Zanini GM, Meays D, et al. (2010) Murine CM is associated with a vasospasm-like microcirculatory dysfunction, and survival upon rescue treatment is markedly increased by nimodipine. *Am J Pathology* 176: 1306-1315.
- 81 Armah H, Doodoo AK, Wiredu EK, et al. (2005) High-level cerebellar expression of cytokines and adhesion molecules in fatal, paediatric, CM. *Ann Trop Med Parasitol* 99: 629-647.
- 82 Favre N, Da Laperousaz C, Ryffel B, et al. (1999) Role of ICAM-1 (CD54) in the development of murine CM. *Micro Infect* 1: 961-968.
- 83 Clark IA, Rockett KA (1994) The cytokine theory of human cerebral malaria. *Parasitol* 10: 410-412.
- 84 Kern P, Hemmer CJ, Van Damme J, et al. (1989) Elevated tumour necrosis factor alpha and interleukin-6 serum levels as markers for complicated *P. falciparum* malaria. *Am J Med* 87: 139-143.
- 85 Lucas R, Lou J, Morel DR (1997) TNF receptors in the microvascular pathology of acute respiratory distress syndrome and CM. *J Leu Biol* 61: 551-558.
- 86 Mackenzie CD, Grau GE, Molyneux ME (1999) Intravascular leucocytes in the brain in Malawian children with fatal malaria. *Am J Trop Med Hyg* 61: 476.
- 87 Isaacs A, Lindenmann J (1957) Virus interference I. The interferon. *Proceedings of the Royal Society of London, UK*.
- 88 Young HA (1996) Regulation of interferon-gamma gene expression. *J Interferon Cytokine Res* 16: 563-568.
- 89 Frucht DM, Fukao T, Bogdan C, et al. (2001) IFN-gamma production by antigen-presenting cells: mechanisms emerge. *Trends Immunol* 22: 556-560.
- 90 Munder M, Mallo M, Eichmann K, (2001) Direct stimulation of macrophages by IL-12 and IL-18-a bridge built on solid ground. *Immunology* 75: 159-160.
- 91 Amani V, Vigário AM, Belnoue E, et al. (2000) Involvement of IFN-g receptor-mediated signalling in pathology and anti-malarial immunity induced by *Plasmodium berghei* infection. *Euro J Immunol* 30: 1646-1655.
- 92 Ho M, Sexton MM, Tongtawe P, et al. (1995) Interleukin-10 inhibits tumour necrosis factor production but not antigen-specific lymphoproliferation in acute *Plasmodium falciparum* malaria. *J Infect Dis* 172: 838-844.
- 93 Ringwald P, Peyron F, Vuillez JP, (1991) Levels of cytokines in plasma during *Plasmodium falciparum* malaria attacks. *J Clin Microb*. 29: 2076-2078.
- 94 Koch O, Awomoyi A, Usen S, (2002) IFNGR1 gene promoter polymorphisms, susceptibility to CM. *J Infect Dis* 185:1684-1687.
- 95 Kishimoto T, Akira S, Taniuchi T (1992) Interleukin-6 and its receptor: a paradigm for cytokines. *Science* 258: 593-597.
- 96 Kern P, Hemmer CJ, Van Damme J, Gruss HJ (1989) Elevated tumor necrosis factor alpha and interleukin-6 serum levels as markers for complicated *P. falciparum* malaria. *Am J Med* 87: 139-143.
- 97 Eling WM, Kreamsner PG (1994) Cytokines in malaria, pathology and protection. *Biotherapy* 7: 211-221.
- 98 Grau GE, Bieler G, Pointaire P, De Kossodo S (1990) Significance of cytokine production and adhesion molecules in malarial immunopathology. *Immunology* 25: 189-194.
- 99 de Kossodo S, Grau GE (1993) Profiles of cytokine production in relation with susceptibility to cerebral malaria. *J Immunol* 151: 4811-4820.
- 100 Donnelly RP, Dickensheets H, Finbloom DS (1999) The interleukin-10 signal transduction pathway and regulation of gene expression in mononuclear phagocytes. *J Inter Cyt Res* 19: 563-573.
- 101 Kossodo S, Monso C, Juillard P, et al. (1997) Interleukin-10 modulates susceptibility in experimental cerebral malaria. *Immunol* 91: 536-540.
- 102 Brown HH (1999) Evidence of blood–brain barrier dysfunction in human CM. *Neuro Appl Neuro* 25: 331-340.
- 103 Grau GE, Taylor TE, Molyneux ME, et al. (1989) Tumour necrosis factor and disease severity in children with falciparum malaria. *N Engl J Med* 320: 1586-1591.
- 104 Moore KW, de Waal Malefyt R, Coffman RL, et al. (2001) Interleukin-10 and the interleukin-10 receptor. *Annu Rev Immunol* 19: 683-765.
- 105 Thumwood CM, Hunt NH, Clark IA, et al. (1988) Breakdown of the blood-brain barrier in murine cerebral malaria. *Parasitology* 96: 579-589.
- 106 Belnoue E, Kayibanda M, Vigario AM, et al. (2002) On the pathogenic role of brain-sequestered alpha CD8+ T cells in experimental cerebral malaria. *J Immunol* 169: 6369-6375.
- 107 Belnoue E, Kayibanda M, Deschemin JC, et al. (2003) CCR5 deficiency decreases susceptibility to experimental cerebral malaria. *Blood* 101: 4253-4259.
- 108 Dinarello CA, Simon A, van der Meer JW (2012) Treating inflammation by blocking interleukin-1 in a broad spectrum of diseases. *N Rev Dis* 11: 633-652.
- 109 Joosten LA (2010) Excessive interleukin-1 signalling determines the development of Th1 and Th17 responses in chronic inflammation. *Arth Rhe* 62: 320-322.
- 110 Curfs JH, van der Meer JW, Sauerwein RW, et al. (1990) Low dosages of interleukin 1 protect mice against lethal cerebral malaria. *J Exp Med* 172: 1287-1291.
- 111 Brown MA, Hural J (1997) Functions of IL-4 and control of its expression. *Crit Rev Immunol* 17: 1-32.
- 112 Von der Weid T, Kopf M, Köhler G, et al. (1994) The immune response to *Plasmodium chabaudi* malaria in interleukin-4-deficient mice. *Eur J Immunol* 24: 2285-2293.
- 113 Kumaratilake LM, Ferrante A (1992) IL-4 inhibits macrophage-mediated killing of *P. falciparum* vitro: a possible parasite-immune evasion mechanism. *J Immunol* 149: 194-199.
- 114 Machado FS, Desruisseaux MS, Nagajyothi, et al. (2006) Endothelin in a murine model of cerebral malaria. *Exp Biol Med* 231: 1176-1181.
- 115 Kennan RP, Machado FS, Lee SC (2005) Reduced cerebral blood flow and N-acetyl aspartate in a murine model of CM. *Parasitol Res* 96: 302-307.
- 116 Dai M, Freeman B, Bruno FP, et al. (2012) The novel ETA receptor antagonist HJP-272 prevents cerebral microvascular haemorrhage

- in CM and synergistically improves survival in combination with an artemisinin derivative. *Life Sci* 91: 687-692.
- 117 McCarron RM, Wang L, Stanimirovic DB, et al. (1993) Endothelin induction of adhesion molecule expression on human brain microvascular endothelial cells. *NeuroLetters* 156: 31-34.
 - 118 Moncada S, Higgs A (1993) The L-arginine-nitric oxide pathway. *N Engl J Med* 329: 2002-2012.
 - 119 Clark IA, Rockett KA, Cowden WB (1991) Proposed link between cytokines, nitric oxide and human cerebral malaria. *Parasitol Today* 7: 205-207.
 - 120 Cabrales P, Zanini GM, Meays D, et al. (2011) Nitric oxide protection against murine CM is associated with improved cerebral microcirculatory physiology. *J Infect Dis*. 203: 1454-1463.
 - 121 Yeo TW, Lampah DA, Gitawati R, et al. (2007) Impaired nitric oxide bioavailability and L-arginine reversible endothelial dysfunction in adults with falciparum malaria. *J Exp Med* 204: 2693-704.
 - 122 Van der Heyde HC, Nolan J, Combes V, et al. (2006) A unified hypothesis for the genesis of CM: sequestration, inflammation and hemostasis leading to microcirculatory dysfunction. *Trends Parasitol* 22: 503-508.
 - 123 Bertinaria M, Guglielmo S, Rolando B, et al. (2011) Amodiaquine analogues containing NO-donor substructures: synthesis and their preliminary evaluation as potential tools in the treatment of CM. *Euro J Med Chem* 46: 1757-1767.
 - 124 Yeo TW, Lampah DA, Tjitra E, et al. (2009) Relationship of cell-free haemoglobin to impaired endothelial nitric oxide bioavailability and perfusion in severe falciparum malaria. *J Inf Dis* 200: 1522-1529.
 - 125 De Macchi BM, Miranda FJ, de Souza FS, et al. (2013) Chickens treated with a nitric oxide inhibitor became more resistant to *P. gallinaceum* infection due to reduced anaemia, thrombocytopenia and inflammation. *Veterin Res* 44.
 - 126 Perkins DJ, Hittner JB, Mwaikambo ED (2005) Impaired systemic production of prostaglandin E2 in children with CM. *J Inf Dis* 191: 1548-1557.
 - 127 Xiao L, Patterson PS, Yang C, et al. (1999) Role of eicosanoids in the pathogenesis of murine cerebral malaria. *Am J Trop Med Hyg* 60: 668-673.
 - 128 Peters-Golden M, Canetti C, Mancuso P, et al. (2005) Leukotrienes: underappreciated mediators of innate immune responses. *J Immunol* 174: 589-594.
 - 129 Peters-Golden M, Henderson WR (2007) Mechanisms of disease: leukotrienes. *N Eng J Med* 357: 1798-1854.
 - 130 Babu KS, Salvi SS (2000) Aspirin and asthma. *Chest* 118: 1470-1476.
 - 131 Arcoleo F, Milano S, D'Agostino P (1995) Effect of exogenous leukotriene B4 (LTB4) on BALB/c mice splenocyte production of Th1 and Th2 lymphokines. *In J of Immun* 17: 457-463.
 - 132 Bate CA, Taverne J, Playfair JH (1988) Malarial parasites induce TNF production by macrophages. *Immunology* 64: 227-231.
 - 133 Kwiatkowski D, Cannon JG, Manogue KR (1989) Tumour necrosis factor production in Falciparum malaria and its association with schizont rupture. *Clin Exp Immunol* 77: 361-366.
 - 134 Schofield L, Hackett F (1993) Signal transduction in host cells by a glycosylphosphatidylinositol toxin of malaria parasites. *J Exp Med* 177: 145-153.
 - 135 Elased KM, Gumaa KA, de Souza JB, et al. (2004) Improvement of glucose homeostasis in obese diabetic db/db mice given *Plasmodium yoelii* glycosylphosphatidylinositols. *Metabolism: Clinl Exp* 53: 1048-1053.
 - 136 Schofield L, Vivas L, Hackett F (1993) Neutralising monoclonal antibodies to glycosylphosphatidylinositol, the dominant TNF-alpha-inducing toxin of *Plasmodium falciparum*: prospects for the immunotherapy of severe malaria. *An Trop Med Para* 87: 617-626.
 - 137 Schofield L, Novakovic S, Gerold P, et al. (1996) Glycosylphosphatidylinositol toxin of *Plasmodium* up-regulates intercellular adhesion molecule-1, vascular cell adhesion molecule-1, and E-selectin expression in vascular endothelial cells and increases.
 - 138 Schwarzer E, Turrini F, Ulliers D, et al. (1992) Impairment of macrophage functions after ingestion of *P. falciparum*-infected erythrocytes or isolated malarial pigment. *J Exp Med* 176: 1033-41.
 - 139 Schwarzer E, De Matteis F, Giribaldi G, et al. (1999) Hemozoin stability and dormant induction of heme oxygenase in hemozoin fed human monocytes. *Mol Bio Parasitol* 100: 61-72.
 - 140 Lyke KE, Diallo DA, Dicko A, et al. (2003) Association of intra-leukocytic *P. falciparum* malaria pigment with disease severity, clinical manifestations and prognosis in severe malaria. *Am J Trop Med Hyg* 69: 253-259.
 - 141 Urban BC, Roberts DJ (2003) Inhibition of T cell function during malaria: implications for immunology and vaccinology. *J Exp Med* 197: 137-141.
 - 142 Novelli EM, Hittner JB (2010) Clinical predictors of severe malarial anaemia in a holoendemic *P. falciparum* transmission area: research paper. *British J Haem* 149: 711-721.
 - 143 Ochiel DO, Awandare GA, Keller CC, Hittner JB (2005) Differential regulation of chemokines in children with *P. falciparum* malaria. *Inf Immun* 73: 4190-4197.
 - 144 Sanni LA, Thomas SR, Tattam B (1998) Dramatic changes in oxidative tryptophan metabolism along the kynurenine pathway in experimental cerebral and nonCM. *Am J Pathol* 152: 611-619.
 - 145 Medana IM, Hien T (2002) The clinical significance of cerebrospinal fluid levels of Kynurenine pathway metabolites and lactate in severe malaria. *J inf Dis* 185: 650-656.
 - 146 Chiarugi A, Dello Sbarba P, Paccagnini A, et al. (2000) Combined inhibition of indoleamine 2,3-dioxygenase and nitric oxide synthase modulates neurotoxin release by interferon-gamma-activated macrophages. *J Leu Biol* 68: 260-266.
 - 147 Clark C, Mackay GM, Smythe (2005) Prolonged survival of a murine model of CM by Kynurenine pathway inhibition. *Inf Immun* 73: 5249-5251.
 - 148 Yeo W, Lampah DA, Gitawati R, et al. (2008) Angiopoietin-2 is associated with decreased endothelial nitric oxide and poor clinical outcome in severe *falciparum* malaria. 102.
 - 149 Lovegrove FE, Tangpukdee N, Opoka RO (2009) Serum angiopoietin-1 and -2 levels discriminate CM from uncomplicated malaria and predict clinical outcome in African children. *PLOS One*. 4: 4912.
 - 150 Taoufiq Z, Gay F, Balvanyos J, Ciceron L (2008) Rho-kinase inhibition in severe malaria: thwarting parasite-induced collateral damage to endothelia. *J Inf Dis* 197: 1062-1073.
 - 151 Zang-Edou ES, Bisvigou U, Taoufiq Z, et al. (2010) Inhibition of *P. falciparum* field isolates-mediated endothelial cell apoptosis by Fasudil: therapeutic implications for severe malaria. *PLoS One* 5: 13221.